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A review of the diversity and impact of invasive non-native species in tropical marine ecosystems

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Abstract

Tropical marine ecosystems are biologically diverse and economically invaluable. However, they are severely threatened from impacts associated with climate change coupled with localized and regional stressors, such as pollution and overfishing. Non-native species (sometimes referred to as ‘alien’ species) are another major threat facing these ecosystems, although rarely discussed and overshadowed by the other stressors mentioned above. NNS can be introduced accidentally (for example via shipping activities) and/or sometimes intentionally (for aquaculture or by hobbyists). Understanding the extent of the impacts NNS have on native flora and fauna often remains challenging, along with ascertaining when the species in question actually became ‘invasive’. Here we review the status of this threat across key tropical marine ecosystems such as coral reefs, algae meadows, mangroves, and seagrass beds. We aim to provide a baseline of where invasive NNS can be found, when they are thought to have been introduced and what impact they are thought to be having on the native ecosystems they now inhabit. In the appended material we provide a comprehensive list of NNS covering key groups such as macroalgae, sponges, seagrasses and mangroves, anthozoans, bryozoans, ascidians, fishes, and crustaceans.

Background

Tropical marine ecosystems (TMEs) are biodiverse and include a variety of habitats, such as coral reefs, mangrove forests, seagrass beds, and algae meadows (Hansen [2003](#)). Collectively TMEs support billions of people worldwide (Kaufman and Dayton [1997](#), Moberg and Folke [1999](#), Barbier et al. [2011](#)), providing ecosystem services, such as coastal defence, fisheries, recreation and tourism, and carbon sequestration (Barbier [2017](#)). Furthermore, coastal TMEs represent an important interface between land and sea, controlling nutrient fluxes and material exchange from terrestrial to open ocean environments (Barbier [2017](#)). However, these habitats are facing unprecedented stressors in the Anthropocene (the epoch associated with humankind) including human mediated climate change as well as local and regional stressors such as over exploitation, coastal development, pollution, and the introduction of non-native species (NNS) (Hansen [2003](#), De Poorter et al. [2009](#)). NNS, which are often referred to alternatively as 'alien', 'non-indigenous', 'introduced' or 'exotic', include any species occurring outside its natural past or present range, dispersed by direct or indirect, intentional, or unintentional human activities (Walther et al. [2009](#)). The term invasive non-native species (or invasive alien) is used when the species becomes established in its new home, spreads outside the introduction area and poses a threat to the native wildlife, the ecosystem functioning, economics, human health and/or society in general (Walther et al. [2009](#), Teixeira and Creed [2020](#)).

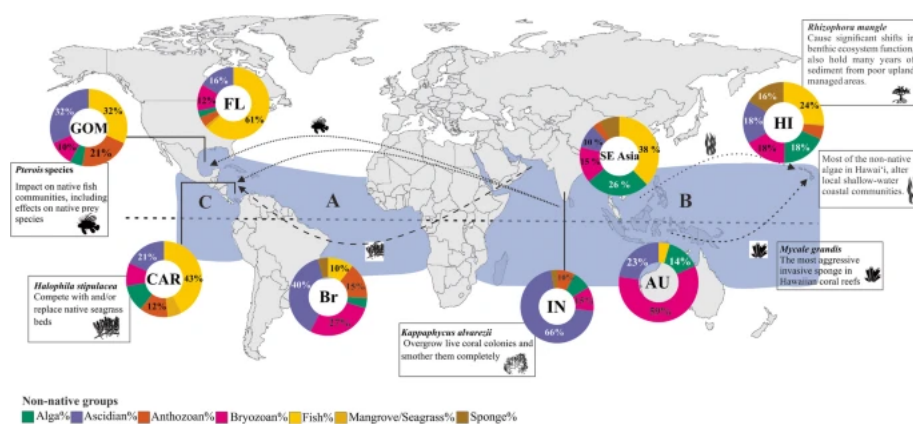
NNS can be introduced and subsequently spread in tropical waters via several different pathways, like any other aquatic environments. However, the relative importance of invasion vectors remains difficult to assess as linking a specific vector with a certain invasion is often impossible. In general, international shipping activities, including ballast water exchanges and vessel fouling, are usually blamed, and are identified as the two primary routes for many species reaching new habitats (Ruiz et al. [1997](#), Godwin [2003](#), Molnar et al. [2008](#)). Aquaculture is likely the next major source of NNS within a region as in some cases escapes of the cultured species may occur from farms. Further, non-target species may inadvertently be introduced as hitch hikers with the imported stock (De Silva et al. [2009](#), Cagauan [2007](#)). Other pathways include introductions from aquaria and zoos or the hobbyist trade, via recreational activities, tourist activities, and transport on marine litter or debris (Bax et al. [2003](#), Padilla and Williams [2004](#), Anderson et al. [2015](#)).

When a NNS is introduced to a new region, there are several obstacles the species need to overcome before it becomes a successful invader. In the tropics, for example, where there is typically high natural biodiversity, complex biotic interactions of the native communities often make it difficult for NNS to get a foothold (Kennedy et al. [2002](#), Kühn and Klotz [2008](#), Wells and Bieler [2020](#)). Although, this is not always the case and some tropical areas have lower than average diversity, Hawaii and the eastern Pacific for example. Regardless, if introduced species do manage to attain high abundances, they have the potential to displace native species, affect ecosystem processes, change community structure, impact human health, decrease native biodiversity and cause substantial economic losses (Mack et al. [2000](#), Grosholz [2002](#), Bax et al. [2003](#), Simberloff [2005](#)). Although not all NNS will have such direct negative impacts in their recipient systems (Simberloff [2005](#), Goodenough [2010](#)), the subsequent introduction of non-native parasites and possibly pathogenic microorganisms which may

occur with any host might be more of a threat (Young et al. [2017](#)). Indeed, this latter aspect of NNS is so poorly understood in the tropics, there are only a few marine examples that can be drawn upon to highlight this impact (see section 5).

Here, we aim to create a baseline data set and review of where NNS can be found with marine ecosystems of tropical regions (including Tropical Atlantic, Indo-Pacific and Eastern Tropical Pacific; Spalding et al. [2007](#)) (Fig. [1](#)). We focused our review on invasive NNS, as by definition these species have much greater impacts on the marine ecosystem they inhabit and are much less numerous than other NNS. Further, due to the large number of documented NNS in general our goal was not to present a complete list of species present, but instead, highlight key tropical marine taxa, such as algae, marine plants (mangroves and seagrass), sessile invertebrates (sponges, bryzoans, ascidians and corals) and fish – and specific examples within these groups. We also attempt to summarize invaded tropical regions, assess introduction pathways for each NNS, explore the threat to native biodiversity and the TMEs where possible (see Additional files [1](#) and [2](#)).

Fig. 1



The review covers tropical marine ecosystems throughout the Tropical Atlantic **a**, Indo-Pacific Ocean **b**, and Eastern Tropical Pacific **c**, (blue colour); in this figure we illustrate the percentage of marine non-native species (NNS) found in different tropical regions (percentages less than 10% are not displayed in the donut charts). Data was generated from the Additional file [1](#). Arrows identify origin of the species now known to be the main invasive NNS in the TMEs which we describe in more detail throughout the review. Key to location names include: Hawai'i (HI), South-East Asia (SE Asia), Australia-tropical waters (AU), India (IN), Brazil-tropical waters (Br), Florida (FL), Caribbean Sea (CAR), and Gulf of Mexico (GOM)

Algae

Marine macroalgae are a major component of NNS globally, with current estimations of introductions in excess of 300 species (Davidson et al. [2015](#)). Many non native (NN) algae, start off in aquaculture or in harbors transported via the ballast water of ships, currents will then often transport the NNS throughout their new habitats (Hewitt et al. [2007](#), Williams and Smith [2007](#), Dubinsky and Stambler [2010](#)). However, this is an over generalization as the pathways of 40% of NN algae globally remains

unknown (Williams and Smith [2007](#)). Here we have identified 57 marine NN algae in tropical regions (Additional file [1](#)). Hawai'i stands out as having the highest number of NN algae (Additional file [2](#)), some of which directly or indirectly threaten local coral reef ecosystems and seagrass beds. Take the red alga *Kappaphycus alvarezii*, for example. This invasive NNS was extensively farmed in the Philippines in the 1960s (Bixler [1996](#), Sulu et al. [2004](#)), in Hawai'i from 1970 (Conklin and Smith [2005](#)) and in the Gulf of Mannar from 1990 (Kamalakkannan et al. [2014](#)). Established populations have since spread outside the farmed area, and it is recognised as an established NNS in India, Tanzania, Panama, Venezuela, Hawai'i and Fiji (Rodgers and Cox [1999](#), Ask et al. [2003](#), Chandrasekaran et al. [2008](#), Sellers et al. [2015a](#)). This destructive tropical red alga is mostly cultured very close to coral ecosystems and its negative impacts have been recorded on coral reefs in many areas, such as Kāne'ōhe Bay in Hawai'i and the Gulf of Mannar, India (Rodgers and Cox [1999](#), Chandrasekaran et al. [2008](#), Patterson Edward and Bhatt [2012](#)). *K. alvarezii* triggers decline in the density and diversity of native fish and brings down the species richness and abundance of native macroalgae, coral and other benthic macrofauna (MoEF&CC Project Report [2018](#), Neilson et al. [2018](#), HISC [2019](#)).

Evidently, the success of *K. alvarezii* is related to its fast growth rate. In Hawai'i, the rate of spread is estimated to be $250 \text{ cm}\cdot\text{year}^{-1}$ with an average growth rate of $5.06\% \text{ day}^{-1}$ (Glenn and Doty [1990](#), HISC [2019](#)). In just over 2 years the alga increased its cover from less than 10% to more than 50% (Conklin and Smith [2005](#)). In the Gulf of Mannar *K. alvarezii* grows over live coral colonies and smothers them completely (Chandrasekaran et al. [2008](#), Patterson Edward and Bhatt [2012](#), Edward et al. [2015](#), MoEF&CC Project Report [2018](#)). This overgrowth can cause a coral-algal phase shift in short periods of time (Conklin and Smith [2005](#), MoEF&CC Project Report [2018](#)), especially when compounded by overfishing of the herbivorous fish species which graze the NN algae (Dubinsky and Stambler [2010](#)). However, even when herbivory was reported as high, *K. alvarezii* appears to be less preferred by many species and so is less grazed (Smith et al. [2002](#), Conklin and Smith [2005](#)).

Interestingly, coral bleaching might, inadvertently, be the answer to controlling *K. alvarezii*, which appears to not be able to survive on dead coral skeleton (MoEF&CC Project Report [2018](#), Barnhill and Bahr [2019](#)). Although we recognize that this is not a 'management strategy', it does offer a window into the future of these ecosystems and phase shifts from coral reef to alga *K. alvarezii* are unlikely to occur with the demise of coral reefs. While phase shifts have been reported in other regions when corals decline in abundance and the ecosystem's herbivores are not present to control algae overgrowth – for example throughout much of the Caribbean (Mumby [2009](#)). In this case, when the corals recover (possibly through extensive restoration practices), there is of course nothing to prevent the remnants of *K. alvarezii* from repopulating and invading the recovering reefs – this will obviously hamper efforts to protect this fragile ecosystem (MoEF&CC Project Report [2018](#), HISC [2019](#)).

Other non-native red algae such as *Gracilaria salicornia*, *Acanthophora spicifera* and *Hypnea musciformis* are also known to cause many problems in Hawai'i. *G. salicornia* was first introduced in

the 1970s for experimental aquaculture for the agar industry (Smith et al. [2002](#), [2004](#)). It quickly monopolizes any available substrate and, in many areas, has become the single-most dominant benthic species in the region. Before the invasion, much higher diversities of algae species were commonly encountered (over 60 species in some examples) – (Smith et al. [2004](#)). This invasive NNS is also able to significantly acidify the water which leads to the decline of coral reefs (Martinez et al. [2012](#)). *A. spicifera* was first detected in Hawai‘i in the 1950s and has since been recorded at Palmyra Atoll in 2002 and 2008 (Russell [1992](#), Russell and Balazs [1994](#), Knapp et al. [2011](#)), whilst *H. musciformis* was first introduced in 1974 for commercial cultivation. These invasive species are frequently observed smothering reef organisms such as corals and native algae (Smith et al. [2002](#)). The rapid dispersal of all three NNS is associated with fast growth rates, morphological plasticity, abundant production of tetraspores (reproduction strategy) and their ability to be epiphytic on other algae (Russell [1992](#), Smith et al. [2002](#)). Interestingly, there may be a glimmer of hope in the form of the green turtle (*Chelonia mydas*). These turtles appear to prefer consumption of NN algae (when given a choice), and so may act as a natural control mechanism preventing the complete overtake of the TMEs by these NNS (Russell and Balazs [1994](#), Russell and Balazs [2015](#)).

Two other species of invasive NN algae, worthy of note are the green algae *Avrainvillea erecta* and *A. amadelpha*, which were ‘accidentally’ introduced to Hawai‘i in the early 1980s. These algae have since become established in sandy and hardbottom habitats which were originally dominated by the endemic Hawaiian sea grass (*Halophila hawaiiiana*), (Martinez et al. [2009](#), Peyton [2009](#), Wade et al. [2018](#)). However, although these NN algal meadows contain approx. 95% more biomass than the seagrass meadows, *H. hawaiiiana* appears the superior competitor, which led researchers to suggest the seagrass would naturally suppress the algae mats preventing complete colonization (Peyton [2009](#)).

Impacts associated with invasive NN algae are generally perceived to be negative, however as the above examples indicate, this can vary and in fact, less than 12% of NN algae are thought to be a major threat to TMEs (Additional files [1](#) and [2](#)). However, this value may likely change once further research monitors the continued spread and succession of these NNS in these habitats.

Marine plants

Mangroves

Native mangrove plants are essential habitat for many marine species in tropical and subtropical regions but predicting and understanding how mangroves become established as NNS and subsequently flourish is challenging. That said, there are several well-documented introductions of mangroves in non-native habitats, which offers us clues about how these species become established over time. For example, humans have intentionally introduced non-native mangroves to both non-native (e.g., Moloka‘i, Hawai‘i) and native (e.g., Bangladesh, China, South Florida, and Tonga) mangrove regions around the world (Fourqurean et al. [2010](#)). In 1902, the red mangrove (*Rhizophora mangle*), originally from South Florida was introduced to Moloka‘i, Hawai‘i with the intent to

stabilize the shoreline (Chimner et al. [2006](#)). Fifteen years later it was introduced to O'ahu for the same purpose (Allen [1998](#)). The red mangrove and the buttonwood (*Conocarpus erectus*), which both originate from South Florida, along with the upriver orange mangrove (*Bruguiera sexangular*), the lenggadai (*Bruguiera parvifolia*), the spurred mangrove (*Ceriops tagal*) and the Asiatic mangrove (*Rhizophora mucronate*), all from the Philippines were later introduced in O'ahu in 1922 (Allen [1998](#)). However, only the red mangrove and the upriver orange mangrove became well established. The red mangrove in particular has successfully invaded all the main Hawaiian Islands except Kaho'olawe and Ni'ihau (Allen [1998](#)). Mangroves are now found on every coast in O'ahu except the dry leeward coast (Allen [1998](#)). Removal projects are underway yet NNS stands still exist, although some populations have been reduced in recent years.

In Miami, Florida, two Indonesian Tumu mangrove trees (*Bruguiera gymnorrhiza*) were planted in the intertidal zone in 1940, and the white-flowered black mangrove (*Lumnitzera racemose*) was introduced multiple times during the mid-1960s and early-1970s (Fourqurean et al. [2010](#)). Today, the Tumu and white-flowered mangrove populations are expanding throughout South Florida at around 5.6 and 17–23% per year, respectively (Fourqurean et al. [2010](#)). They can compete with native species because their natural environments (Indo-Pacific), life-history characteristics, and genetics are similar to native mangroves, including the frequency of disturbance at the introduction sites (Fourqurean et al. [2010](#)).

Non-native mangroves in general can alter chemical and physical conditions and cause numerous problems in regions where they do not naturally occur like Hawai'i for example. The red mangrove causes obnoxious odors (anoxic conditions) and clogs tidal streams (Chimner et al. [2006](#)). Non-native mangroves can also cause significant shifts in benthic ecosystem function, with sediment metabolism, benthic community structure and short-term C-remineralization dynamics being affected for up to 6-years following invader removal (Sweetman et al. [2010](#)). Non-native mangroves can also restrict tidal flow, and lower dissolved oxygen concentrations (Allen [1998](#)); low flow and anoxic conditions can also cause toxic algal blooms (Chimner et al. [2006](#)). Moreover, mangrove roots can trap fine and organic-rich sediment, which alters benthic communities and has long-term and broad implications for native marine fauna (Demopoulos and Smith [2010](#)). Also, the red mangrove can negatively impact native biodiversity, especially in coastal fish ponds and habitat quality for endangered shorebirds, such as the Hawaiian stilt (*Himantopus mexicanus knudseni*) and the Hawaiian moorhen (*Gallinula chloropus sanvicensis*) (Allen [1998](#), Chimner et al. [2006](#)). Non-native mangroves are not only fast growing, but they can crowd out native species. In Hawai'i, all mangroves are non-native and have colonized many different coastal habitats, including tidal flats, riverbanks, fishponds, canals, protected reefs, embayments, lagoons, and other protected areas (Chimner et al. [2006](#)). It should be noted that there is debate on the removal of them in some areas. While they may impact native biodiversity and alter native shorelines, they are also holding many years of sediment from poorly managed upland areas, so removing them may increase sediment onto the reefs offshore. Previous removal efforts have focused in specific areas where managers have the ability to control the

population. However, there is no data to demonstrate the positive or negative impacts of their removal in these areas.

Mangrove ecosystems themselves often resist invasion of other NNS, because the harsh saline conditions do not allow most other invasive plants and trees to establish (Lugo [1998](#)). However, non-native mangroves in Hawai'i have facilitated the establishment of other NNS, such as the Samoan crab (*Scylla serrata*), barnacles (e.g., *Chthamalus proteus*, *Amphibalanus reticulatus*, and *Amphibalanus amphitrite*) and sponges (e.g., *Suberites aurantiacus*, *Haliclona (soestella) caerulea*, and *Gelloides fibrosa*) (Eldredge and Smith [2001](#), Demopoulos and Smith [2010](#)). Impacts associated with the introduction of non-native mangrove plants are generally perceived to be negative, but some studies suggest positive impacts through the increase in the abundance and diversity of species occurring in an area (Demopoulos et al. [2007](#)).

Seagrass

Seagrasses can rapidly grow in new areas through vegetative propagation, altering the seascape, and transforming soft sandy bottoms into areas of submerged aquatic vegetation (Steiner and Willette [2015](#), Smulders et al. [2017](#)). Initially monospecific patches will form but with high growth rates, meadows soon proliferate if given a chance (Willette and Ambrose [2009](#)). *Halophila stipulacea*'s native range is from India to the eastern continent of Africa, Madagascar, the Red Sea and the Persian Gulf (Den Hartog [1970](#), Short et al. [2007](#)). This species is often referred to as a 'lessepsian migrant' (i.e. invaded the Mediterranean Sea through the Suez Canal, which opened in 1869) (Gambi et al. [2009](#)). From the canal it spread west to Malta, the Ionian Sea and northern Sicily (Willette and Ambrose [2009](#)). In 2002 for the first time the occurrence of this NNS was documented in the tropical waters off the Caribbean coast of Grenada, West Indies (Willette and Ambrose [2009](#), Willette et al. [2014](#), Ruiz et al. [2017](#)), and subsequently in the British Virgin Islands and Puerto Rico (Ruiz et al. [2017](#)) and west to Aruba and the Venezuelan mainland (Vera et al. [2014](#), Willette et al. [2014](#)). This species has extended its range dramatically, roughly 700 km over 15 years (Chiquillo et al. [2019](#)). Spread is likely facilitated by boats (on anchors or intertwined with fishing traps) and/or more 'naturally' via ocean currents (Ruiz and Ballantine [2004](#), Willette and Ambrose [2012](#), Willette et al. [2014](#), Ruiz et al. [2017](#)). Until recently it was thought that Caribbean populations disseminated exclusively by fragments of natural and anthropogenic origin, yet flowers and fruits have since been recorded in Caribbean populations throwing doubt onto this hypothesis (Vera et al. [2014](#), Chiquillo et al. [2019](#), Willette et al. [2020](#)).

H. stipulacea competes with native seagrass often replacing species such as *Syringodium filiforme*, *Halodule wrightii*, *H. decipiens* and even the climax species *Thalassia testudinum* (Willette and Ambrose [2012](#), Steiner and Willette [2015](#), Smulders et al. [2017](#), Christianen et al. [2019](#)). Although *H. stipulacea* sometimes presents a sparse growth form, which does not overgrow the local seagrasses, it can often form an understory layer preventing other species propagating themselves (Van Tussenbroek et al. [2016](#)). *H. stipulacea* is also highly resilient to small-scale disturbance and may grow into damaged areas 1.5–30 times faster than native seagrasses (Willette et al. [2020](#)). It is therefore likely to thrive under situations of anthropogenic impact such as poorly flushed Caribbean bays, which are often

subjected to high nutrient inputs and physical damage in harbours and anchorages (Olinger et al. [2017](#)). Moreover, grazers such as green turtle (*Chelonia mydas*) may modify the rate and spatial extent of this invasive species' expansion, due to grazing preferences, and increased space for settlement (Becking et al. [2014](#), Christianen et al. [2019](#)).

Canopy heights of *H. stipulacea* are much lower than native seagrasses (Olinger et al. [2017](#)) so it also alters the abundance and composition of seagrass-associated organisms, with knock-on effects at different trophic levels (Willette and Ambrose [2009](#), Boman et al. [2019](#)). For example, *H. stipulacea* patches have been documented to harbour reduced fish diversity and alter juvenile fish assemblages, with an abundance of some nocturnal carnivores and scarcity of herbivores and diurnal carnivores (Olinger et al. [2017](#)).

H. stipulacea is an exceptional ecologically flexible invasive NNS and although it can provide new seagrass habitat (Willette and Ambrose [2012](#), Pinault et al. [2018](#)), its continued expansion (Rogers et al. [2014](#)) may: 1) compromise seagrass ecosystem functioning (Smulders et al. [2017](#)); and 2) negatively interact with *H. engelmanni* and *H. baillonii*, both of which have small ranges and are listed as “vulnerable” by the IUCN (Short et al. [2011](#), Willette et al. [2014](#)).

Native seagrass communities are also almost certainly affected by several other invasive NNS (Hansen [2003](#)), yet there is little literature on any specific impacts. In this regard, Williams ([2007](#)) listed 56 known NNS in seagrass beds. Of those only four macroalgae (*K. alvarezii*, *Eucheuma denticulatum*, *A. amadelpha* and *Caulerpa ollivieri*) have been introduced into tropical seagrass meadows. Furthermore, the Indo-Pacific lionfish *Pterois volitans* (Claydon et al. [2012](#)) and Indo-Pacific green mussel *Perna viridis* (Buddo et al. [2003](#)) have been reported in tropical Caribbean seagrass beds. The invasive swimming crab *Charybdis hellerii*, native to the Indo-West Pacific, occurs almost exclusively in dense *H. stipulacea* beds rather than sparse ones, native seagrass beds, sand, rocks or algae (Ferry et al. [2017](#)).

Sessile invertebrates

Sponges

In the context of NNS, sponges have received little attention in TMEs, and their impacts remain largely unknown. However, it does appear that invasive NN sponges are able to compete for space with native sponges and corals, thereby posing a possible threat to reef ecosystems as their health declines due to climate change (Eldredge and Smith [2001](#)). The transfer of NN sponges is likely accidental, through shipping, commercial trawling, and marine aquaculture (Godwin [2003](#), Carballo et al. [2013](#), Klautau et al. [2020](#)). According to our database, 34 marine NN sponges have been documented in tropical regions (with the highest numbers again reported from Hawai'i), (Additional files [1](#) and [2](#)). For example, the orange keyhole sponge (*Mycale grandis*), native to Indonesia, was unintentionally introduced to Hawai'i in 1996 (Coles et al. [2007](#), Coles and Bolick [2007](#)). Unlike the majority of other NN sponges found in Hawai'i, this aggressive alien sponge has been described as a

major threat to corals as the observed long-term sustained biomass implies adaptability of this species to future climate change scenarios/environments (Coles et al. [2007](#), Coles and Bolick [2007](#), Shih [2018](#)). This invasive NNS is able to withstand high pCO₂ and high temperatures which have proven stressful for many other reef organisms (Vicente et al. [2016](#)). Biomass of *M. grandis* increased throughout the mid-2000s and the sponge remains a prominent member of the benthic community in Kāneʻohe Bay and other partially degraded shallow coral reef ecosystems in Hawaiʻi (Shih [2018](#)). Large clusters of this species are often present in shaded areas such as along pier pilings and docks, but it also grows in clear and shallow locations on the reef.

In the Mexican Pacific, another NN sponge, *Chalinula nematifera* (originally native of the Indo-Pacific) was first identified in 2003 on coral reefs in the Isla Isabel National Park. *C. nematifera* overgrows corals, as well as dead skeletons and coralline algae (Ávila and Carballo [2009](#), Turicchia et al. [2018](#)). This species grows at a rapid rate (50 mm.mo⁻¹), which is comparatively higher than most of the native sponges in the Mexican Pacific. With such a rapid rate of spread, there is the high potential for this sponge to dominate coral reefs in these regions (Rossi et al. [2015](#), Turicchia et al. [2018](#)).

The other NN sponges we were able to find evidence of (Additional file [1](#)) are even less well understood than the above examples. One explanation for this may be the lack of studies which even explore these species in their traditional home ranges, those throughout the Indo-Pacific in this instance (Van Soest et al. [2012](#), Carlton and Eldredge [2015](#)). Further, we can also not rule out that some of these 'NNS' may not be non-native, as the invasion pathways and origin of many of them are not clear to date (see Additional files [1](#) and [2](#)).

Bryozoans and ascidians

Fouling organisms such as bryozoans and ascidians (tunicates) frequently constitute the majority of NNS recorded, as they attach regularly to any natural and man-made structures (McCann et al. [2007](#), Shenkar and Swalla [2011](#)). Shipping is again thought to be one of the major pathways for their introductions (Lambert and Lambert [1998](#), Lambert [2002](#), see Additional file [2](#)). These organisms are also known to spread through aquaculture (i.e., transport of contaminated shells or shellfish between mariculture operation), rafting of floating debris carried by ocean currents, and attached to live species (Lambert [2001](#), Bernier et al. [2009](#), Tamilselvi et al. [2011](#)). For example, 49 species of Western Pacific coastal bryozoans were found on 317 floating objects that originated from the Great East Japan Earthquake and Tsunami of 2011 (McCuller and Carlton [2018](#)). These objects (and their hitchhikers) drifted across the North Pacific Ocean and landed in Hawaiʻi and North America (McCuller and Carlton [2018](#)). As the majority of the drifting objects were plastics, one can therefore classify the species carried upon them as true NNS (Stelfox et al. [2020](#)).

These fouling organisms dominate ports and harbors (Marins et al. [2010](#), Sheets et al. [2016](#)), and their impacts therefore have a direct economical aspect (Eldredge and Smith [2001](#), Lambert [2002](#), Miranda et al. [2018c](#)). They have also been shown to decrease species richness in general, act as

habitat modifiers, impact on many epifaunal species and change biodiversity by overgrowing and competing with native species (Wallentinus and Nyberg [2007](#), Lutz-Collins et al. [2009](#), Shenkar and Swalla [2011](#)). NN tunicates for example compete with several commercially important aquaculture species, such as mussels. This is done via competition for food (plankton in the water column) and/or occupation of the settlement space (Tamilselvi et al. [2011](#), Zhang et al. [2019](#)). However, healthy natural ecosystems such as coral reefs (often characterised by high biodiversity) appear able to resist the establishment of NN ascidians (Kennedy et al. [2002](#), Lambert [2002](#)). In Guam, for example, most NN ascidians are confined to harbor structures and have not yet significantly colonized natural reef tracts (Lambert [2002](#)).

We found 125 NN bryozoans and ascidians documented in shallow waters and coastal areas (Additional file [1](#)). However, owing to the lack of systematic and biogeographical data on native species and natural diversity of these groups the number of NNS is likely to be an underestimate. Some species have wide geographical distributions, and it is impossible to rule out historical introductions before baseline studies assessed their home ranges. Take *Amathia verticillata* for example (one of the most widely recorded NNS in the world; Marchini et al. [2015](#), Minchin et al. [2016](#), McCann et al. [2015](#), Collins et al. [2020](#)). Such a cosmopolitan distribution has posed questions as to its real native origin and many are arguing that despite first being described in Italy, its origin may have been the Caribbean (Galil and Gevili [2014](#), Marchini et al. [2015](#), Miranda et al. [2018c](#)). *A. verticillata*'s economic impact comes from its extensive fouling behaviour on harbors and marinas, as well as on the vessels and fishing gear, and it has been known to clog water intake pipes (Miranda et al. [2018c](#)). Also, this NNS is known to cause local biodiversity impacts due to its dominance over native species (Miranda et al. [2018c](#)). Furthermore, several studies have highlighted that other fauna (such as bryozoans, nudibranchs, amphipods and isopods) are associated with this bryozoan. *A. verticillata* itself is therefore likely a special niche for smaller NNS, enhancing opportunities for their spread as well (Farrapeira [2011](#), Marchini et al. [2015](#)).

Although NN bryozoans and ascidians are common and diverse, they do appear largely restricted to harbors and ports, struggling to gain a foothold in natural ecosystems (Lambert [2001](#), Ali et al. [2009](#)). For this reason, they do have an economic impact which warrants attention, however their ecological impact in TMEs may be much less than other groups of NNS. However, there is no guarantee that this will remain as such as these systems change and transgress in a changing world.

Hard and soft corals

Although not the first thought when discussing NNS, there are several classic coral examples which we can draw on here (see Additional file [1](#)), such as the three scleractinians *Tubastrea coccinea*, *T. tagusensis*, and *T. micranthus*. Of these, *T. coccinea* and *T. tagusensis* are considered highly invasive NNS and cause significant environmental, economic, and social impacts as they spread in Brazil (Creed [2006](#), Lages et al. [2011](#), Mantelatto and Creed [2015](#)). *T. micranthus* has a similar potential for negative impacts although less studied to date (Sammarco et al. [2010](#), [2013](#), [2014](#)). Interestingly, *T. coccinea* and *T. tagusensis* co-occur in many areas, growing on top of each other and often coalescing (Creed et

al. [2017b](#)). They often exclude native corals (Creed [2006](#), dos Santos et al. [2013](#), Miranda et al. [2016b](#)) and other invertebrates (Silva et al. [2019](#)) where they grow, modifying coral recruitment and fish assemblages (Miranda et al. [2018a, b](#)), transforming communities and seascapes (Lages et al. [2011](#), De Paula et al. [2017](#), Capel et al. [2020](#)), and impacting ecosystem service provision (Mantelatto and Creed [2015](#), De Paula et al. [2017](#)). They are proven effective invaders as they exploit multiple reproductive modes and are fast growing (Mizrahi et al. [2014](#), De Paula et al. [2014](#), Capel et al. [2017](#), Luz et al. [2018](#)). They also employ physical and chemical defenses against competitors and predators (Lages et al. [2010ab](#), Lages et al. [2012](#), dos Santos et al. [2013](#)).

Originally from the Indo-Pacific, these species are widely found throughout the tropical and sub-tropical Atlantic including Brazil, throughout the Caribbean Sea, the Gulf of Mexico (GOM), and the Canary Islands (Creed et al. [2017a](#), López et al. [2019](#)). *T. coccinea* and *T. micranthus* naturally occur in the Indo-Pacific (Cairns and Zibrowius [1997](#), Sammarco et al. [2013](#)), whilst *T. tagusensis* was originally restricted to the Galapagos Archipelago (Ecuador) (Wells [1982](#), Glynn and Wellington [1983](#)).

T. coccinea was first reported as a NNS in 1943 in Aruba and Puerto Rico (Western Atlantic) (Vaughan and Wells [1943](#), Fenner and Banks [2004](#)). Currently, its non-native range extends to the Caribbean Sea, the GOM (Fenner [2001](#)), Florida (Fenner and Banks [2004](#)) and Georgia (Creed et al. [2017a](#)), the southwestern Atlantic (late 1980s) (Fenner [1999](#), De Paula and Creed [2004](#), Miranda et al. [2016a](#), de Faria and Kitahara [2020](#)), and recently the Canary Islands (López et al. [2019](#)).

T. tagusensis similarly has been reported as cryptogenic in the Persian Gulf (Creed et al. [2017a](#)), non-native in the Brazilian southwest Atlantic (De Paula and Creed [2004](#), Mantelatto et al. [2011](#), Sampaio et al. [2012](#), Creed et al. [2017a](#), de Oliveira Soares et al. [2018](#)), in GOM on artificial reefs and oil platforms (Figueroa et al. [2019](#)), and in the Canary Islands (López et al. [2019](#)). *T. tagusensis* has likely been in the Caribbean and GOM for > 15 years and historically overlooked (Joel Creed pers. obs. in photographs and T. Shearer, pers. communication). Therefore, it is possible that it arrived together with *T. coccinea* prior to 1943 in the Atlantic and in the 1980s and 2010s in Brazil and the Canary Islands, respectively.

The third species, *T. micranthus*, is thus far only found on oil and gas platforms operating in the (extra-tropical) northern GOM - with reports of colonies at depths ≤ 183 m (Sammarco et al. [2010](#), [2013](#), [2014](#)). Interestingly, a fourth, yet unidentified clade of *Tubastraea* (with intermediate characteristics) has also been recognized in the GOM (Figueroa et al. [2019](#)) and Brazil (Joel C. Creed, pers. obs.), but additional taxonomic work needs to be completed to determine its status or significance – note the whole genus likely needs revision (Capel et al. [2019](#)).

Creed et al. ([2017a](#)) reviewed pathways and vectors of *Tubastraea* spp. and concluded that their introduction to the Caribbean and GOM was from fouling on ships' hulls, especially floating platforms, and their current distribution is also a result of passive dispersal of larvae by currents. In the southwest Atlantic, Gabon, and the Canary Islands, *Tubastraea* spp. were likely introduced

through biofouling on oil platforms and/or drill ships, probably redeployed from western Africa, the GOM, or other areas throughout the Indo-Pacific (Friedlander et al. [2014](#), López et al. [2019](#), Creed et al. [2017a](#)). Except for the Caribbean, the earliest records of *Tubastraea* in the Atlantic are on oil platforms and genetic studies have further confirmed that oil platforms serve as the most likely vector (Capel et al. [2019](#)).

In addition to *Tubastraea* other scleractinians have also become NNS. For example, the mushroom coral *Lobactis scutaria* (previously known as *Fungia scutaria*) is native to the Indo- and Central Pacific and Red Sea, but in 1966 it was introduced to Discovery Bay, Jamaica from Eilat, Israel. Specimens were held on the reef for laboratory-based experiments at the Discovery Bay Marine Laboratory where they remained until Thomas F. Gorwau (the researcher working on them) died in 1970 (Bush et al. [2004](#)). Over the next 10 years, several attempts to eradicate this species were made with 25 adult individuals removed (LaJeunesse et al. [2005](#)), but two additional individuals were found in 2003 (Bush et al. [2004](#)). The fact that the species survived so long was considered by Bush et al. ([2004](#)) “as an ominous warning of potential future invasions by other tropical marine species”. The ecological impact however is thought to be minor for this NNS and its successful eradication may have now occurred with the removal of these remaining two colonies (LaJeunesse et al. [2005](#)). This example is unusual in that it was introduced by scientists. Similarly, *Siderastrea siderea* was introduced into Pacific Panama as five skeleton blocks thought to be dead and used in an experiment, but then discovered to be living and named *Siderastrea glynni* by Budd and Guzman (1994). All known colonies were collected and are kept in aquaria (Glynn et al. [2016](#)).

There are also several soft corals that have been documented as NNS in TMEs. For example, seven octocorals including three species of Clavulariidae, two species of Xeniidae, one species of Anthothelidae and one species of Nephtheidae are considered NNS in the tropics. All three clavulariids have been introduced into the southwest Atlantic (Brazil). While *Carijoa riseii* is a widespread invasive fouling species (see below), *Clavularia* cf. *viridis* has only been reported from one location as an aquarium release (Mantelatto et al. [2018](#)) and has since been removed. Another clavulariid that presents an interesting example is *Stragulum bicolor*. This soft coral is classified as a type 3 pseudoindigenous species (sensu Carlton [2009](#)), i.e. a species described as ‘new’ after introduction and remaining unknown elsewhere). Currently this species is only found in Brazil. Its highly conspicuous appearance in areas previously surveyed and its ability to colonize artificial substrates (van Ofwegen and Haddad [2011](#)) is evidence that it has been introduced, although it remains unclear where it may have originated from. However, the presence of this species appears to have little effect on the native community dynamics (Altvater and Coutinho [2015](#)).

The nephtheid *Chromonephthea braziliensis* is another example of a type 3 pseudoindigenous species (sensu Carlton [2009](#)). Again, first described from Brazil, it was documented to have been introduced at Cabo Frio, Rio de Janeiro, in the mid-1990s (van Ofwegen [2005](#), Ferreira et al. [2009](#)). Compared to the native soft corals, it is a very large and distinctive species, with high dominance in certain sites. Similar to *Tubastraea*, it was most likely introduced by oil platforms and drill ships that are known to

anchor at the initial site of discovery (Ferreira et al. [2006](#)). In 1 year, a single colony has been reported to increase to 40 individuals (Ferreira et al. [2009](#)) and the species is expanding its range slowly (12 km in ≈ 25 years). It has been shown to cause tissue mortality in the native endemic gorgonian *Phyllogorgia dilatata* (Lages et al. [2006](#), Fleury et al. [2008](#)), although the extent of the ecological impacts remain unclear.

In 2017, another type 3 pseudoindigenous species (sensu Carlton [2009](#)) was identified, this time a xeniid from the genus *Sansibia*. This soft coral was first detected on the shallow subtidal tropical rocky reefs in southeast Brazil (Mantelatto et al. [2018](#)). To date, attempts at describing the species (via morphological and genetic analyses) have not been able to place this as one known to science (Mantelatto et al. [2018](#)) – note the genus was only described in 2000. Interestingly, *Sansibia* was associated positively with some macroalgae and negatively with the zoantharian *Palythoa caribaeorum*, which probably provided greater biotic resistance to invasion (Mantelatto et al. [2018](#)). This NNS was discovered at the same time and same site as both *C. viridis* (see above) and *Erythropodium caribaeorum* (Anthothelidae) – (Carpinelli et al. [2020](#)). As all three are typical ornamental species kept by hobbyists, they were likely introduced through this route. This undescribed *Sansibia* sp. is currently estimated at expanding at a rate of 40 m yr^{-1} – assuming a linear and constant expansion rate (Mantelatto et al. [2018](#)).

Moving to Venezuela now, *Xenia membranacea* was identified as a NNS when a single colony appears in 2007 on the coastal coral community of Valle Seco (Allais et al. [2014](#)). Originally from Indonesia this species has now increased in abundance, dominates substrate, overgrows native corals, and has extended its range by several kilometers. As this invasive NNS appears to have severe impacts on the native fauna and overgrows on scleractinian corals such as *Colpophyllia natans* and *Montastraea cavernosa*, this is certainly a species to watch in the coming years (Allais et al. [2014](#)).

The snowflake coral, *Carijoa riisei*, is another interesting example of a NNS in this group. *C. riisei* was thought to have originated from the western Atlantic (Carlton and Eldredge [2009](#)), but is now believed to be native to the Indo-Pacific (Concepcion et al. [2010](#)). It is now known to have a wide distribution across the Indo-Pacific and western Atlantic; however, its original native distribution is not known, again due to taxonomic uncertainty. In Hawai'i it was first found in 1966 (Concepcion et al. [2010](#)) and formally recognised as a NNS in this region in 1972 (Evans et al. [1974](#)). It can currently be found on all the Main Hawaiian Islands (Carlton and Eldredge [2009](#), Montgomery A. pers. obs.). While Carlton and Eldredge ([2009](#)) reported *C. riisei* to be introduced in Hawai'i, Concepcion et al. ([2010](#)) concluded the origin of the Hawai'i colonies remains unknown, so there is an argument that *C. riisei* should be classified as cryptogenic for the Hawaiian Archipelago. Based on genetic analysis by Concepcion et al. ([2010](#)) it may be more likely that western Atlantic populations were introduced from the Pacific than vice versa.

C. riisei is now reported as a NNS in India – in the Gulf of Mannar (Padmakumar et al. [2011](#), Raghunathan et al. [2013](#)) and as a potential NNS in the Galapagos (Collins et al. [2020](#)). However, this

determination is not based on evidence of its prior absence. In Colombia, large, dense stands of *C. riisei* are recorded but appear sporadically (Sánchez and Ballesteros [2014](#)). In the Eastern Pacific, *Carijoa* grows on gorgonians instead of the black corals it grows on in Hawai'i (*Antipathes griggi* and *A. grandis*). Another genetic study explored the origin of these octocorals and indicated that these Eastern Pacific populations had close affinities to Western Atlantic populations suggesting they arrived into the Eastern Pacific relatively recently, probably by ship through the Panama Canal (Quintanilla et al. [2017](#)). The rise of notoriety of *C. riisei* in the Hawaiian Islands specifically was based on the species ability to overgrow native black coral colonies (Grigg [2003](#)). Grigg ([2003](#)) reported that depths between 75 and 110 m were a “virtual graveyard for black corals”. Subsequent quantitative analysis of this area showed approximately 60% of the colonies between 80 and 100 m had been colonized by this NNS (Kahng and Grigg [2005](#)). *C. riisei* appears to favour the larger black coral colonies and appears to initially attach to dead branches, then smother the living tissue (Kahng and Grigg [2005](#)). Interestingly, results from a survey conducted in 2006 indicated that the ‘invasion’ appears to be abating (Kahng [2007](#)), so the current threat of the NNS remains unknown.

Fishes

Invasions by fishes, especially those occupying higher trophic levels, are much rarer in comparison to marine plants or invertebrates (Byrnes et al. [2007](#), Côté et al. [2013](#)). Although the rate of introduction of fishes may be initially low, once they have been introduced they have a high potential (~ 50%) of becoming established, much higher than their plant counterparts for example (Jeschke and Strayer [2005](#)). Fishes that are successful in establishing can, and usually do, have severe impacts on their invaded ecosystem through predation, most notably piscivory, which can drive population dynamics and structure the rest of the community (Hixon [2015](#)). Invasive predators are also likely to have important non-consumptive ‘fear’ effects (Mitchell and Harborne [2020](#)), but these have rarely been investigated for tropical marine species (see Kindinger [2015](#), Eaton et al. [2016](#), Kindinger and Albins [2017](#) for exceptions). While several fishes have been introduced accidentally (via global trade or the aquaculture industry), others have been introduced intentionally by governmental agencies, most often with the purpose of opening new fisheries to local communities or enhance existing ones (Randall [1987](#)) (see Additional file [1](#)).

For the purpose of this review, we are only focusing on established NNS, but recognize that individuals of many other species have been observed in NN regions without becoming established (e.g. see Semmens et al. [2004](#), and Schofield and Akins [2019](#), for fish species seen in Florida). NN fish species that are not fully marine (brackish, anadromous) such as members of the family Cichlidae or Salmonidae that have been intentionally introduced globally (De Silva [2004](#), Buoro et al. [2016](#)) are also excluded from this list. However, it should be noted that many of these species have drastic impacts on native flora/fauna and some of them are described as vectors of diseases and parasites (See Section 5). Additionally, for some species it is unclear if their non-native presence is due to introduction or is simply a natural range expansion of nearby native populations (e.g. *Gramma loreto*

in Florida and *Hypsoblennius invemar* in Brazil). With the above caveats, we will focus on examples of non-native fish species that are invasive and/or those which have invasive characteristics.

Perhaps the best studied marine vertebrate NNS is the Indo-Pacific lionfish (*Pterois* spp.), and its invasion in the western Atlantic. Lionfish were first observed off the coast of Florida in 1985 (Schofield [2009](#)), with a likely introduction of a few individuals from the aquarium trade (Whitfield et al. [2002](#), Betancur-R et al. [2011](#)). Sightings remained low for several years until the first report of lionfish in the Bahamas occurred in 2004. This was followed by a rapid increase in abundance and range expansion throughout the Caribbean to South America (Schofield [2009](#)). Currently, the entire non-native distribution of lionfish spans as far south as Venezuela east to Bermuda, throughout the GOM, and year-round northern populations persist along the coast of North Carolina (Schofield [2010](#), Côté and Smith [2018](#)). The expansion of the known range of this fish in the western Atlantic can be seen at <https://nas.er.usgs.gov/taxgroup/fish/Lionfishanimation.gif>. While the northwards range expansion of the lionfish in the western Atlantic may be near their thermal physiological limits (Kimball et al. [2004](#)), their expansive invase range here and their new invasion in the Mediterranean Sea indicates that TMEs are not the only ecosystems being impacted by this NNS (Kletou et al. [2016](#)).

There are several studies which report the negative impacts of lionfish. However, there are an equal number of contrasting studies which suggest the impact is not as clear cut as one would imagine. For example, in the Bahamas and Florida there are reports of significant declines in native prey populations (Albins and Hixon [2008](#), Green et al. [2012](#)), declines in overall species richness (Albins [2013](#), Albins and Hixon [2013](#), Albins [2015](#), Ellis and Faletti [2016](#)), and local extinctions of a few native species (Ingeman [2016](#)). However, in contrast, in Venezuela and Belize, no such changes in native species abundance or richness were observed (Elise et al. [2015](#), Hackerott et al. [2017](#)). In the southeast US, the decline of adult tomtate populations (*Haemulon aurolineatum*) has also been attributed to lionfish predating on their juvenile population (Ballew et al. [2016](#)). However, other studies again find no such trends (i.e. no decline in specific fish populations due to losses of early life stages) (Green and Côté [2014](#), Albins [2015](#)). The same appears true with competition driven by this new predator. Some studies have shown dramatic declines in native meso-predator populations (Lesser and Slattery [2011](#)) and altered behavior of native species (Henderson [2012](#), Raymond et al. [2015](#)), whilst others show no such trends (Elise et al. [2015](#), Ballew et al. [2016](#)). In fact, some find that native meso-predators may actually reduce the negative impacts of the invasive alien (Ellis and Faletti [2016](#)). Such conflicting data suggests lionfish impact native fish populations in complex speciespecific and/or geographically distinct ways. However, the invasion may be entering a new phase in the western Atlantic. An ulcerative skin condition appears to be plaguing lionfish in their new range (Harris et al. [2020](#)) and their densities appear to be dropping in some locations (Benkwitt et al. [2017](#)).

Another NNS, the regal demoiselle (*Neopomacentrus cyanomos*) is a more recent case of invasion. Originally from the Indo-Pacific, *N. cyanomos* was reported in the GOM, near the reefs of Coatzacoalcos, Mexico in 2013 (González-Gándara and de la Cruz-Francisco [2014](#)). Since then, it was documented to rapidly spread throughout the northern coastal region of the GOM (Robertson et al.

[2016](#), Bennett et al. [2019](#)). The front of the non-native region is currently near the Florida Panhandle just south of Panama City Beach (USGS NAS). In addition to the expanding distribution in the GOM, a second site of introduction was documented in Trinidad in 2019 (Robertson et al. [2021](#)). Given the locations of introduction, the current distribution, and the density of populations at specific locations, it is hypothesized that *N. cyanomos* arrived to the Atlantic as hitchhikers on oil platforms, like how the coral *Tubastraea micranthus* was thought to be introduced into Brazil (Robertson et al. [2016](#), Robertson et al. [2021](#)). The impacts of this NNS remain unknown, but visual observations from divers suggest that *N. cyanomos* potentially competes with and displaces native planktivores such as *Chromis multilineata* (González-Gándara and de la Cruz-Francisco [2014](#), Robertson et al. [2016](#)).

The muzzled blenny *Omobranchus punctatus*, is another good example of a vertebrate NNS. Native to the Indo-Pacific, it has spread through the Caribbean and Brazil since the 1930s (Gerhardinger et al. [2006](#), Lasso-Alcalá et al. [2011](#), Soares et al. [2011](#)). The proximity of most records to ports, indicate that this NNS was initially introduced to the Atlantic by ships travelling from India to Trinidad and then into Brazil by shipping associated with oil platforms (Lasso-Alcalá et al. [2011](#)) – a similar theme to the introductions reported above.

Although never intended for introduction into natural habitats, incidences of non-native fish species escaping mariculture facilities are an all too common occurrence and a major source of NNS. In Singapore for example, non-native fishes procured by the Agri-Food and Veterinary Authority (now Singapore Food Authority) such as *Chanos chanos*, *Sciaenops ocellatus*, and *Larimichthys crocea* have been recorded as escapees and established populations off the coast (Jaafar et al. [2012](#)). The cobia, *Rachycentron canadum*, is yet another example. This circum-global tropical and sub-tropical species is a highly valued food fish reared in floating cages worldwide (Benetti et al. [2010](#)). In 2015, cages were installed off the coast of Ecuador, a region lacking native populations. Several months after, the Ecuadorian Ministry of Environment reported that individuals escaped from one of these cages. Local fishermen have reported catching some of these escapees (Castellanos-Galindo et al. [2016](#)). This NNS has since been discovered off the coasts of Colombia, Peru, and Panama, within only a few months after the initial recorded escape (Vega et al. [2016](#)). The potential impacts of this species on the native biota are yet unexplored. However, this predatory NNS reaches sexual maturity early, is highly fecund, and resides in a variety of coastal habitats, suggesting a high potential to disrupt the native TMEs of the Tropical Eastern Pacific (Castellanos-Galindo et al. [2018](#)).

The opening of the Panama Canal in 1914 is also worthy of briefly mentioning here as it is a major pathway for NNS invasions (Gunter [1979](#), Cohen [2006](#)). While most of these NNS are confined to the lakes within the canal system (McCosker and Dawson [1975](#)), and will therefore not be discussed in this review, there is one NN fish that crossed from the Atlantic to the Pacific. The Atlantic tarpon (*Megalops atlanticus*) now has stable populations in the Tropical Eastern Pacific, with an expanding distribution. Hildebrand ([1937](#)) first described their presence in the lakes near the Pacific entrance of the canal in the 1930s. This was followed by Swanson's ([1946](#)) observation of tarpon outside the entrance. Currently their distribution in the Pacific ranges from Guatemala to the border of Colombia

and Ecuador (Cohen [2006](#), Castellanos-Galindo et al. [2019](#)). While the impacts of the tarpon introduction to the Tropical Eastern Pacific are unknown, their trophic status suggests potential negative effects on native prey communities (Castellanos-Galindo et al. [2019](#)).

Although many of the previous examples were the results of accidental introductions, intentional introductions have also occurred in several cases. In the 1950s, the Division of Fish and Game of the State of Hawai‘i intentionally introduced several marine fish species into the wild either to increase the diversity of local reef fish communities or to open new fisheries. One such event was the introduction of *Sardinella marquesensis* from the Marquesas Islands to O‘ahu in 1955, initially intended to provide bait for tuna. Although established, the abundance of *S. marquesensis* remains relatively low compared to native and other invasive baitfish species (see below; Randall [1987](#)). In addition, the vessels that transported *S. marquesensis* to O‘ahu unintentionally contained several other species, including two NNS (*Osteomugil engeli* -previously known as *Valamugil engeli*- and *Upeneus vittatus*; Randall [1987](#), Mundy [2005](#)). Both have since become established, but only *O. engeli* has populations in high abundance with the potential to compete with native species, such as the commercially important mullet species *Mugil cephalus* (Randall [1987](#)). Interestingly another NNS (*Herklotsichthys quadrimaculatus*), which was also unintentionally introduced to Hawai‘i in 1975 (likely brought over by tuna fishing vessels returning from the Marshall Islands) (Baldwin [1984](#), Randall [1987](#), Mundy [2005](#)) appears to have been the cause for the low abundances of *S. marquesensis* (as well as other native baitfish species).

Another event of intentional introduction in Hawai‘i included members of the families Lutjanidae, Lethrinidae, and Serranidae. In the 1950s, eleven species were released and three became established (Randall [1987](#), Mundy [2005](#)): *Cephalopholis argus* and *Lutjanus fulvus* from Moorea and *Lutjanus kasmira* from the Marquesas. Unlike *L. fulvus* populations that remain relatively low, *L. kasmira* is highly abundant. The latter species has invaded the entire Hawaiian archipelago, all islands in both the Main Hawaiian Islands and the Northwestern Hawaiian Islands. It is perceived to reduce catches of more valuable native species and potentially compete with and displace native species (Randall [1987](#), Oda and Parrish [1981](#), Friedlander et al. [2002](#), Schumacher and Parrish [2005](#)). That said, *C. argus* has received more research attention than the two snapper species due to possible human health concerns. After the initial release of this NNS, there was a considerable lag time before reaching substantial populations in the 1980s (Crooks [2005](#)). Once populations were high, there was renewed interest in establishing a fishery around this species. That was until reports of ciguatera poisoning in humans arose after their consumption (Dierking and Campora [2009](#)). Despite only 18.2% of *C. argus* individuals having toxins high enough to be harmful to humans (Dierking and Campora [2009](#)), the concern of poisoning has essentially closed the fishery. Populations of this piscivorous species are now unchecked and have the potential to impact the native prey communities (Dierking et al. [2009](#)).

Non-native diseases in tropical marine ecosystems

Disease-causing organisms (parasites and pathogens) are common in all ecosystems and cause a plethora of diseases in various species (Groner et al. [2016](#), Harvell [2019](#)). Introductions of NNS has been accredited as being a major way to either directly or indirectly modulate the normal interplay between parasites and their hosts, and could serve as one of the main drivers of novel emerging diseases in TMEs, ultimately affecting biodiversity, ecosystem function and ecosystem services/human health (Crowl et al. [2008](#), Goedknecht et al. [2016](#); see also Torchin et al. [2002](#), Blakeslee et al. [2013](#)). In this final section of the review, we exclusively focus on diseases that have been shown to be caused by NNS (either through introducing a NN host or a NN parasite). For the sake of this review, we are defining parasites as ‘organisms living on or in another organism, deriving substances from the host organism, without benefit to the host’. We are therefore using this broad definition including parasitoids and other pathogens (disease-causing organisms) that ultimately produce damage in the host (Prenter et al. [2004](#)).

Any NNS, when first introduced would always have the potential of carrying its natural parasite load with it. During the ‘invasion’ process, this load could be released or significantly reduced in number (Lymbery et al. [2014](#), Goedknecht et al. [2016](#)). However, if only a few of these parasites make the journey with the host NNS, they are likely to parasitise new hosts which would likely be more susceptible to these new disease causing organisms (Vignon and Sasal [2010](#), Lymbery et al. [2014](#)). Farmed escapees are likely the main group of NNS which will carry and spread the highest number of their parasites into any new range. For instance, NN shrimps brought to a given country for aquaculture practices (such as *Penaeus monodon* and *Penaeus vannamei*) have been known to carry several viral pathogens such as monodon baculovirus (MBV), infectious hypodermal and hematopoietic necrosis virus (IHHNV), hepatopancreatic parvovirus (HPV), and taura syndrome virus (TSV) (O’Connor et al. [2008](#), Hernández [2020](#); for more detail about shrimp diseases see Briggs et al. [2004](#) and Sweet and Bateman [2016](#)). It has been hypothesized that these viruses could easily transmit to native shrimp populations, especially if the aquaculture farms are ‘open’ to the sea (Briggs et al. [2004](#), Walker and Mohan [2009](#), Sweet and Bateman [2016](#)).

In other cases, a NNS can exhibit a depauperate ensemble of parasites in its introduced region. For example, the coexistence of gill ectoparasites (such as copepods and monogeneans) have been found to be associated with the peacock grouper, *Cephalopholis argus*, in its native range (Lo et al. [2001](#)) while comparative parasitological studies have shown that *C. argus* loses a significant amount of its native metazoan parasite community when introduced into new areas (Hawai’i for example, see section 4) (Vignon et al. [2009](#)).

NNS may also play a role as reservoirs for native parasites, increasing their abundance and possible impact (Prenter et al. [2004](#)). That said, these native parasites may well limit the demographic performance of a NNS (Sellers et al. [2015b](#), Goedknecht et al. [2016](#)). For example, as discussed above the with regard to the lionfish (*Pterois* spp.), they now appear to be acquiring new parasites within their invaded range (see section 4). In this example, this NNS may be more susceptible to parasites due to its unnaturally high population densities and low genetic diversity – all common traits in the

NNS discussed throughout this review. Ulcerative Skin Disease was only reported to affect lionfish in 2017 and may be why we are starting to see declines in this NNS across parts of its invaded range (Harris et al. [2018](#), [2020](#)). That said, there is also another negative impact this host-parasite interaction may yield, i.e. that of parasite spillback. This is when native parasites cause mass infections in an NNS, and then subsequently the parasite transmits back to the native host population causing an increase in disease amongst native hosts (Harris et al. [2020](#)).

Alternatively if invasive parasites are co-introduced with their invasive host and then subsequently transmit to local native species this is referred to as parasite spill-over (Goedknecht et al. [2016](#)). The bluestripe snapper (*Lutjanus kasmira*) (see section 4) has been shown to carry an intestinal nematode (*Spirocamallanus istiblenni*), which can 'spill over' to native Hawaiian fishes for example (Gaither et al. [2013](#)).

It is also possible for NN parasites to arrive independent of their host. This is primarily via a vector such as ballast water (Goedknecht et al. [2016](#)). It is very difficult to identify when an ecosystem is invaded by a NN parasite species, but the rapid regional spread of a newly identified disease within a specific group of animals could highlight the likelihood of this happening. An example is the mass die-off of the tropical sea urchin *Diadema antillarum* in the Western Atlantic (Lessios et al. [1984](#), Lessios [1988](#)). A similar disease, this time affecting *Echinothrix calimaris* and *Diadema paucispinum*, also occurred in Hawaii around the same time (1981). In both these cases, the causal agent or agents was not known, yet the disease/s was thought to originate near the Caribbean entrance to the Panama Canal suggesting both might be related or even the same (Birkeland [1989](#)).

Another example is the newly observed coral disease in Florida, dubbed 'stony coral tissue loss disease' (or SCTLTD). This disease impacts many coral species and is particularly virulent, spreading north and south of its reported origin (Florida) (Aeby et al. [2019](#), Precht et al. [2016](#)) and is now being reported across much of the Caribbean. Ballast water has again been highlighted as a possible cause of the rapid spread (U.S. Coast Guard [2019](#), Sweet et al. [2021](#)). There is therefore real concern that the parasite(s) (again as of yet unknown origin), could be transported through the Panama Canal from the Atlantic and into the Indo-Pacific in the not-so-distant future. If such a hypothesis came true, this would be a clear example of the high potential for many more parasites to warrant classification as NNS.

Although it is likely that every NNS carries with it some form of parasite, information on this topic remains scarce. This is primarily due to the difficulties in detecting and identifying these organisms. Further, many parasites are likely to be cryptogenic and their impact would almost always go undetected in short-term studies (Gaither et al. [2013](#), Lohan et al. [2020](#)). This is certainly an important area for future research and will be paramount in assessing the true impact NNS have on their new habitats. Further, survival of these NN parasites will likely be dependant on the survival and success of the original host, the host specificity of the parasite and the ability (or not) of the parasite to produce a resting or dormant stage (see Torchin et al. [2002](#), Lohan et al. [2020](#)).

Conclusion

Non-native species are an integral component of marine tropical ecosystems. NNS have been introduced to the tropical waters, deliberately and incidentally, and some of them have become established, invaded other areas, and caused detrimental ecological impacts, examples of which we give in this review. Such impacts mostly occur through competition, predation, habitat alteration, and disease. However, knowledge of the extent of impacts NNS have on native biota and ecosystems remain incomplete (e.g. see Lesser and Slattery [2011](#)). This is primarily because many NNS are inconspicuous or difficult to identify or locate.

It was evident when compiling this review that Hawai'i stood out as being heavily impacted by NNS (see Additional file [2](#)). Whether this is because of its location, governance or the level of research undertaken to understand NNS in this region we can not tease apart. One factor may be the lower native biodiversity associated with Hawaiian ecosystems (Hutchings et al. [2002](#)). Indeed, areas with higher biodiversity have been shown to be less susceptible (in general) to invasions by NNS (Kühn and Klotz [2008](#), Wells and Bieler [2020](#)). This is primarily due to fewer vacant niches being available to the NNS (Hewitt [2002](#), Kennedy et al. [2002](#)). Therefore, we may very well witness a rise in the number of NNS which become established in TMEs when the results of regional anthropogenic impacts and global climate change occur i.e. declines in the native flora and fauna (Occhipinti-Ambrogi [2007](#), Worm and Lotze [2016](#)). Therefore, there is an urgent need to first gain a better understanding of the species already present in any given TME i.e. a detailed baseline survey conducted across space and time. This will allow us to detect when a new NNS is introduced to a given area and explore invasion pathways more reliably. Armed with this knowledge we may even be able to mitigate and manage the impacts from newly emerging NNS before they become established and uncontrollable. Although several countries are indeed taking an active stance in creating inventories of NNS, many (especially in the Indo-Pacific) are less diligent.

Availability of data and materials

All data generated or analysed during this study are included in this published article [supplementary information: Additional files [1](#) and [2](#)].

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Supplementary Information

[Additional file 1.](#)

A list of marine non-native species in tropical regions (including Tropical Atlantic, Indo-Pacific and Tropical Eastern Pacific).

[Additional file 2.](#)

Reference list for Additional file [1](#), plus additional table and figures.

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